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U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

09/807261

INTERNATIONAL APPLICATION NO.

PCT/US99/24331

INTERNATIONAL FILING DATE

19 October 1999

PRIORITY DATE CLAIMED

19 October 1998

TITLE OF INVENTION

Method of Diagnosing, Monitoring, Staging, Imaging and Treating Prostate Cancer

APPLICANT(S) FOR DO/EO/US

SALCEDA, Susana et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
- a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
- b. ☐ has been transmitted by the International Bureau.
- c. ☒ is not required, as the application was filed in the United States Receiving Office (RO/US).
- ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
- ☒ A copy of the International Search Report (PCT/ISA/210).
- ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
- a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
- b. ☐ have been transmitted by the International Bureau.
- c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
- d. ☒ have not been made and will not be made.
- ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
- ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)). - **unexecuted**
- ☒ A copy of the International Preliminary Examination Report (PCT/PEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

Items 13 to 20 below concern document(s) or information included:

13. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ Certificate of Mailing by Express Mail
20. ☒ Other items or information:

- 1) Courtesy copy of International Application;
2) Copy of Written Opinion; and
3) Return Post Card

"Express Mail" Label No. EL718096860US

Date of Deposit April 10, 2001

I hereby certify that this paper is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Assistant Commissioner for Patents, Box PCT, Washington, D.C. 20231.

By

Typed Name: Deborah Ehret

1 0 APR 2001

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.53) 09/807201		INTERNATIONAL APPLICATION NO. PCT/US99/24331		ATTORNEY'S DOCKET NUMBER DEX-0196	
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21. The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :				CALCULATIONS PTO USE ONLY	
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1,000.00					
<input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00					
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00					
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$860.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	12 - 20 =	0	x \$18.00	\$0.00	
Independent claims	7 - 3 =	4	x \$80.00	\$320.00	
Multiple Dependent Claims (check if applicable) <input type="checkbox"/>				\$0.00	
TOTAL OF ABOVE CALCULATIONS =				\$1,180.00	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable). <input type="checkbox"/>				\$0.00	
SUBTOTAL =				\$1,180.00	
Processing fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30 +				\$0.00	
TOTAL NATIONAL FEE =				\$1,180.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/>				\$0.00	
TOTAL FEES ENCLOSED =				\$1,180.00	
				Amount to be refunded	\$
				charged	\$

☐ A check in the amount of _____ to cover the above fees is enclosed.
☒ **Credit card Payment form for \$1,180.00**
☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees.
 A duplicate copy of this sheet is enclosed.

☒ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. **50-1619** A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

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April 10, 2001
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METHOD OF DIAGNOSING,
MONITORING, STAGING, IMAGING AND TREATING PROSTATE CANCER

FIELD OF THE INVENTION

This invention relates, in part, to newly developed
5 assays for detecting, diagnosing, monitoring, staging,
prognosticating, imaging and treating cancers, particularly
prostate cancer.

BACKGROUND OF THE INVENTION

Cancer of the prostate is the most prevalent malignancy
10 in adult males, excluding skin cancer, and is an increasingly
prevalent health problem in the United States. In 1996, it
was estimated that 41,400 deaths would result from this
disease in the United States alone, indicating that prostate
cancer is second only to lung cancer as the most common cause
15 of death in the same population. If diagnosed and treated
early, when the cancer is still confined to the prostate, the
chances of cure is significantly higher.

Treatment decisions for an individual are linked to the
stage of prostate cancer present in that individual. A common
20 classification of the spread of prostate cancer was developed
by the American Urological Association (AUA). The AUA system
divides prostate tumors into four stages, A to D. Stage A,
microscopic cancer within prostate, is further subdivided into
stages A1 and A2. Sub-stage A1 is a well-differentiated
25 cancer confined to one site within the prostate. Treatment
is generally observation, radical prostatectomy, or radiation.
Sub-stage A2 is a moderately to poorly differentiated cancer
at multiple sites within the prostate. Treatment is radical
prostatectomy or radiation. Stage B, palpable lump within the
30 prostate, is also further subdivided into sub-stages B1 and
B2. In sub-stage B1, the cancer forms a small nodule in one

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lobe of the prostate. In sub-stage B2, the cancer forms large or multiple nodules, or occurs in both lobes of the prostate. Treatment for sub-stages B1 and B2 is either radical prostatectomy or radiation. Stage C is a large cancer mass involving most or all of the prostate and is also further subdivided into two sub-stages. In sub-stage C1, the cancer forms a continuous mass that may have extended beyond the prostate. In sub-stage C2, the cancer forms a continuous mass that invades the surrounding tissue. Treatment for both these sub-stages is radiation with or without drugs to address the cancer. The fourth stage, Stage D is metastatic cancer and is also subdivided into two sub-stages. In sub-stage D1, the cancer appears in the lymph nodes of the pelvis. In sub-stage D2, the cancer involves tissues beyond lymph nodes. Treatment for both of these sub-stages is systemic drugs to address the cancer as well as pain.

However, current prostate cancer staging methods are limited. As many as 50% of prostate cancers initially staged as A2, B, or C are actually stage D, metastatic. Discovery of metastasis is significant because patients with metastatic cancers have a poorer prognosis and require significantly different therapy than those with localized cancers. The five year survival rates for patients with localized and metastatic prostate cancers are 93% and 29%, respectively.

Accordingly, there is a great need for more sensitive and accurate methods for the staging of a cancer in a human to determine whether or not such cancer has metastasized and for monitoring the progress of a cancer in a human which has not metastasized for the onset of metastasis.

It has now been found that a number of proteins in the public domain are useful as diagnostic markers for prostate cancer. These diagnostic markers are referred to herein as cancer specific genes or CSGs and include, but are not limited to: Pro109 which is a human zinc- α 2-glycoprotein (Freje et al. Genomics 1993 18(3):575-587); Pro112 which is a human

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cysteine-rich protein with a zinc-finger motif (Liebhaber et al. Nucleic Acid Research 1990 18(13):3871-3879; WO9514772 and WO9845436); Proll11 which is a prostate-specific transglutaminase (Dubbink et al. Genomics 1998 51(3):434-444);

5 Proll15 which is a novel serine protease with transmembrane, LDLR, and SRCR domains and maps to 21q22.3 (Paoloni-Giacobino et al. Genomics 1997 44(3):309-320; WO9837418 and WO987093); Proll10 which is a human breast carcinoma fatty acid synthase (U.S. Patent 5,665,874 and WO9403599); Proll13 which is a

10 homeobox gene, HOXB13 (Steinicki et al. J. Invest. Dermatol. 1998 111:57-63); Proll14 which is a human tetraspan NET-1 (WO9839446); and Proll18 which is a human JM27 protein (WO9845435). ESTs for these CSGs are set forth in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13 and 15 while the full length contigs for

15 these CSGs are set forth in SEQ ID NO:2, 4, 6, 8, 10, 12, 14 and 16, respectively. Additional CSGs for use in the present invention are depicted herein in SEQ ID NO: 17, 18, 19 and 20.

In the present invention, methods are provided for detecting, diagnosing, monitoring, staging, prognosticating,

20 imaging and treating prostate cancer via the cancer specific genes referred to herein as CSGs. For purposes of the present invention, CSG refers, among other things, to native protein expressed by the gene comprising a polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15,

25 16, 17, 18, 19 or 20. By "CSG" it is also meant herein polynucleotides which, due to degeneracy in genetic coding, comprise variations in nucleotide sequence as compared to SEQ ID NO: 1-20, but which still encode the same protein. In the alternative, what is meant by CSG as used herein, means the

30 native mRNA encoded by the gene comprising the polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20, levels of the gene comprising the polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or

35 20, or levels of a polynucleotide which is capable of

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hybridizing under stringent conditions to the antisense sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20.

Other objects, features, advantages and aspects of the present invention will become apparent to those of skill in the art from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

SUMMARY OF THE INVENTION

Toward these ends, and others, it is an object of the present invention to provide a method for diagnosing the presence of prostate cancer by analyzing for changes in levels of CSG in cells, tissues or bodily fluids compared with levels of CSG in preferably the same cells, tissues, or bodily fluid type of a normal human control, wherein a change in levels of CSG in the patient versus the normal human control is associated with prostate cancer.

Further provided is a method of diagnosing metastatic prostate cancer in a patient having prostate cancer which is not known to have metastasized by identifying a human patient suspected of having prostate cancer that has metastasized; analyzing a sample of cells, tissues, or bodily fluid from such patient for CSG; comparing the CSG levels in such cells, tissues, or bodily fluid with levels of CSG in preferably the same cells, tissues, or bodily fluid type of a normal human control, wherein an increase in CSG levels in the patient versus the normal human control is associated with prostate cancer which has metastasized.

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Also provided by the invention is a method of staging prostate cancer in a human which has such cancer by identifying a human patient having such cancer; analyzing a sample of cells, tissues, or bodily fluid from such patient for CSG; comparing CSG levels in such cells, tissues, or bodily fluid with levels of CSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in CSG levels in the patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of CSG is associated with a cancer which is regressing or in remission.

Further provided is a method of monitoring prostate cancer in a human having such cancer for the onset of metastasis. The method comprises identifying a human patient having such cancer that is not known to have metastasized; periodically analyzing a sample of cells, tissues, or bodily fluid from such patient for CSG; comparing the CSG levels in such cells, tissue, or bodily fluid with levels of CSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in CSG levels in the patient versus the normal human control is associated with a cancer which has metastasized.

Further provided is a method of monitoring the change in stage of prostate cancer in a human having such cancer by looking at levels of CSG in a human having such cancer. The method comprises identifying a human patient having such cancer; periodically analyzing a sample of cells, tissues, or bodily fluid from such patient for CSG; comparing the CSG levels in such cells, tissue, or bodily fluid with levels of CSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in CSG levels in the patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of CSG is associated with a cancer which is regressing or in remission.

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Further provided are methods of designing new therapeutic agents targeted to a CSG for use in imaging and treating prostate cancer. For example, in one embodiment, therapeutic agents such as antibodies targeted against CSG or fragments of such antibodies can be used to detect or image localization of CSG in a patient for the purpose of detecting or diagnosing a disease or condition. Such antibodies can be polyclonal, monoclonal, or omniclonal or prepared by molecular biology techniques. The term "antibody", as used herein and throughout the instant specification is also meant to include aptamers and single-stranded oligonucleotides such as those derived from an *in vitro* evolution protocol referred to as SELEX and well known to those skilled in the art. Antibodies can be labeled with a variety of detectable labels including, but not limited to, radioisotopes and paramagnetic metals. Therapeutics agents such as antibodies or fragments thereof can also be used in the treatment of diseases characterized by expression of CSG. In these applications, the antibody can be used without or with derivatization to a cytotoxic agent such as a radioisotope, enzyme, toxin, drug or a prodrug.

Other objects, features, advantages and aspects of the present invention will become apparent to those of skill in the art from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to diagnostic assays and methods, both quantitative and qualitative for detecting, diagnosing, monitoring, staging and prognosticating cancers

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by comparing levels of CSG in a human patient with those of CSG in a normal human control. For purposes of the present invention, what is meant by CSG levels is, among other things, native protein expressed by the gene comprising a polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20. By "CSG" it is also meant herein polynucleotides which, due to degeneracy in genetic coding, comprise variations in nucleotide sequence as compared to SEQ ID NO: 1-20, but which still encode the same protein. The native protein being detected, may be whole, a breakdown product, a complex of molecules or chemically modified. In the alternative, what is meant by CSG as used herein, means the native mRNA encoded by the gene comprising the polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20, levels of the gene comprising the polynucleotide sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20, or levels of a polynucleotide which is capable of hybridizing under stringent conditions to the antisense sequence of SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20. Such levels are preferably determined in at least one of, cells, tissues and/or bodily fluids, including determination of normal and abnormal levels. Thus, for instance, a diagnostic assay in accordance with the invention for diagnosing overexpression of CSG protein compared to normal control bodily fluids, cells, or tissue samples may be used to diagnose the presence of prostate cancer.

All the methods of the present invention may optionally include determining the levels of other cancer markers as well as CSG. Other cancer markers, in addition to CSG, useful in the present invention will depend on the cancer being tested and are known to those of skill in the art.

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Diagnostic Assays

The present invention provides methods for diagnosing the presence of prostate cancer by analyzing for changes in levels of CSG in cells, tissues or bodily fluids compared with levels of CSG in cells, tissues or bodily fluids of preferably the same type from a normal human control, wherein an increase in levels of CSG in the patient versus the normal human control is associated with the presence of prostate cancer.

Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating the patient being tested has cancer is one in which cells, tissues or bodily fluid levels of the cancer marker, such as CSG, are at least two times higher, and most preferably are at least five times higher, than in preferably the same cells, tissues or bodily fluid of a normal human control.

The present invention also provides a method of diagnosing metastatic prostate cancer in a patient having prostate cancer which has not yet metastasized for the onset of metastasis. In the method of the present invention, a human cancer patient suspected of having prostate cancer which may have metastasized (but which was not previously known to have metastasized) is identified. This is accomplished by a variety of means known to those of skill in the art.

In the present invention, determining the presence of CSG levels in cells, tissues or bodily fluid, is particularly useful for discriminating between prostate cancer which has not metastasized and prostate cancer which has metastasized. Existing techniques have difficulty discriminating between prostate cancer which has metastasized and prostate cancer which has not metastasized and proper treatment selection is often dependent upon such knowledge.

In the present invention, the cancer marker levels measured in such cells, tissues or bodily fluid is CSG, and are compared with levels of CSG in preferably the same cells, tissue or bodily fluid type of a normal human control. That

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is, if the cancer marker being observed is just CSG in serum, this level is preferably compared with the level of CSG in serum of a normal human control. An increase in the CSG in the patient versus the normal human control is associated with prostate cancer which has metastasized.

Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating the cancer in the patient being tested or monitored has metastasized is one in which cells, tissues or bodily fluid levels of the cancer marker, such as CSG, are at least two times higher, and most preferably are at least five times higher, than in preferably the same cells, tissues or bodily fluid of a normal patient.

Normal human control as used herein includes a human patient without cancer and/or non cancerous samples from the patient; in the methods for diagnosing or monitoring for metastasis, normal human control may preferably also include samples from a human patient that is determined by reliable methods to have prostate cancer which has not metastasized.

Staging

The invention also provides a method of staging prostate cancer in a human patient. The method comprises identifying a human patient having such cancer and analyzing cells, tissues or bodily fluid from such human patient for CSG. The CSG levels determined in the patient are then compared with levels of CSG in preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in CSG levels in the human patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of CSG (but still increased over true normal levels) is associated with a cancer which is regressing or in remission.

Monitoring

Further provided is a method of monitoring prostate cancer in a human patient having such cancer for the onset of

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metastasis. The method comprises identifying a human patient having such cancer that is not known to have metastasized; periodically analyzing cells, tissues or bodily fluid from such human patient for CSG; and comparing the CSG levels determined in the human patient with levels of CSG in preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in CSG levels in the human patient versus the normal human control is associated with a cancer which has metastasized. In this method, normal human control samples may also include prior patient samples.

Further provided by this invention is a method of monitoring the change in stage of prostate cancer in a human patient having such cancer. The method comprises identifying a human patient having such cancer; periodically analyzing cells, tissues or bodily fluid from such human patient for CSG; and comparing the CSG levels determined in the human patient with levels of CSG in preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in CSG levels in the human patient versus the normal human control is associated with a cancer which is progressing in stage and a decrease in the levels of CSG is associated with a cancer which is regressing in stage or in remission. In this method, normal human control samples may also include prior patient samples.

Monitoring a patient for onset of metastasis is periodic and preferably done on a quarterly basis. However, this may be more or less frequent depending on the cancer, the particular patient, and the stage of the cancer.

Assay Techniques

Assay techniques that can be used to determine levels of gene expression (including protein levels), such as CSG of the present invention, in a sample derived from a patient are well known to those of skill in the art. Such assay methods include, without limitation, radioimmunoassays, reverse transcriptase PCR (RT-PCR) assays, immunohistochemistry

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assays, *in situ* hybridization assays, competitive-binding assays, Western Blot analyses, ELISA assays and proteomic approaches: two-dimensional gel electrophoresis (2D electrophoresis) and non-gel based approaches such as mass spectrometry or protein interaction profiling. Among these, ELISAs are frequently preferred to diagnose a gene's expressed protein in biological fluids.

An ELISA assay initially comprises preparing an antibody, if not readily available from a commercial source, specific to CSG, preferably a monoclonal antibody. In addition a reporter antibody generally is prepared which binds specifically to CSG. The reporter antibody is attached to a detectable reagent such as radioactive, fluorescent or enzymatic reagent, for example horseradish peroxidase enzyme or alkaline phosphatase.

To carry out the ELISA, antibody specific to CSG is incubated on a solid support, e.g. a polystyrene dish, that binds the antibody. Any free protein binding sites on the dish are then covered by incubating with a non-specific protein such as bovine serum albumin. Next, the sample to be analyzed is incubated in the dish, during which time CSG binds to the specific antibody attached to the polystyrene dish. Unbound sample is washed out with buffer. A reporter antibody specifically directed to CSG and linked to a detectable reagent such as horseradish peroxidase is placed in the dish resulting in binding of the reporter antibody to any monoclonal antibody bound to CSG. Unattached reporter antibody is then washed out. Reagents for peroxidase activity, including a colorimetric substrate are then added to the dish. Immobilized peroxidase, linked to CSG antibodies, produces a colored reaction product. The amount of color developed in a given time period is proportional to the amount of CSG protein present in the sample. Quantitative results typically are obtained by reference to a standard curve.

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A competition assay can also be employed wherein antibodies specific to CSG are attached to a solid support and labeled CSG and a sample derived from the host are passed over the solid support. The amount of label detected which is attached to the solid support can be correlated to a quantity of CSG in the sample.

Nucleic acid methods can also be used to detect CSG mRNA as a marker for prostate cancer. Polymerase chain reaction (PCR) and other nucleic acid methods, such as ligase chain reaction (LCR) and nucleic acid sequence based amplification (NASABA), can be used to detect malignant cells for diagnosis and monitoring of various malignancies. For example, reverse-transcriptase PCR (RT-PCR) is a powerful technique which can be used to detect the presence of a specific mRNA population in a complex mixture of thousands of other mRNA species. In RT-PCR, an mRNA species is first reverse transcribed to complementary DNA (cDNA) with use of the enzyme reverse transcriptase; the cDNA is then amplified as in a standard PCR reaction. RT-PCR can thus reveal by amplification the presence of a single species of mRNA. Accordingly, if the mRNA is highly specific for the cell that produces it, RT-PCR can be used to identify the presence of a specific type of cell.

Hybridization to clones or oligonucleotides arrayed on a solid support (i.e. gridding) can be used to both detect the expression of and quantitate the level of expression of that gene. In this approach, a cDNA encoding the CSG gene is fixed to a substrate. The substrate may be of any suitable type including but not limited to glass, nitrocellulose, nylon or plastic. At least a portion of the DNA encoding the CSG gene is attached to the substrate and then incubated with the analyte, which may be RNA or a complementary DNA (cDNA) copy of the RNA, isolated from the tissue of interest. Hybridization between the substrate bound DNA and the analyte can be detected and quantitated by several means including but

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not limited to radioactive labeling or fluorescence labeling of the analyte or a secondary molecule designed to detect the hybrid. Quantitation of the level of gene expression can be done by comparison of the intensity of the signal from the analyte compared with that determined from known standards. The standards can be obtained by *in vitro* transcription of the target gene, quantitating the yield, and then using that material to generate a standard curve.

Of the proteomic approaches, 2D electrophoresis is a technique well known to those in the art. Isolation of individual proteins from a sample such as serum is accomplished using sequential separation of proteins by different characteristics usually on polyacrylamide gels. First, proteins are separated by size using an electric current. The current acts uniformly on all proteins, so smaller proteins move farther on the gel than larger proteins. The second dimension applies a current perpendicular to the first and separates proteins not on the basis of size but on the specific electric charge carried by each protein. Since no two proteins with different sequences are identical on the basis of both size and charge, the result of a 2D separation is a square gel in which each protein occupies a unique spot. Analysis of the spots with chemical or antibody probes, or subsequent protein microsequencing can reveal the relative abundance of a given protein and the identity of the proteins in the sample.

The above tests can be carried out on samples derived from a variety of cells, bodily fluids and/or tissue extracts such as homogenates or solubilized tissue obtained from a patient. Tissue extracts are obtained routinely from tissue biopsy and autopsy material. Bodily fluids useful in the present invention include blood, urine, saliva or any other bodily secretion or derivative thereof. By blood it is meant to include whole blood, plasma, serum or any derivative of blood.

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In Vivo Targeting of CSGs

Identification of these CSGs is also useful in the rational design of new therapeutics for imaging and treating cancers, and in particular prostate cancer. For example, in one embodiment, antibodies which specifically bind to CSG can be raised and used *in vivo* in patients suspected of suffering from prostate cancer. Antibodies which specifically bind a CSG can be injected into a patient suspected of having prostate cancer for diagnostic and/or therapeutic purposes.

The preparation and use of antibodies for *in vivo* diagnosis is well known in the art. For example, antibody-chelators labeled with Indium-111 have been described for use in the radioimmunoscentigraphic imaging of carcinoembryonic antigen expressing tumors (Sumerdon et al. Nucl. Med. Biol. 1990

17:247-254). In particular, these antibody-chelators have been used in detecting tumors in patients suspected of having recurrent colorectal cancer (Griffin et al. J. Clin. Onc. 1991 9:631-640). Antibodies with paramagnetic ions as labels for use in magnetic resonance imaging have also been described

(Laufer, R.B. Magnetic Resonance in Medicine 1991 22:339-342). Antibodies directed against CSG can be used in a similar manner. Labeled antibodies which specifically bind CSG can be injected into patients suspected of having prostate cancer for the purpose of diagnosing or staging of the disease

status of the patient. The label used will be selected in accordance with the imaging modality to be used. For example, radioactive labels such as Indium-111, Technetium-99m or Iodine-131 can be used for planar scans or single photon emission computed tomography (SPECT). Positron emitting

labels such as Fluorine-19 can be used in positron emission tomography. Paramagnetic ions such as Gadolinium (III) or Manganese (II) can be used in magnetic resonance imaging (MRI). Localization of the label permits determination of the spread of the cancer. The amount of label within an organ or

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tissue also allows determination of the presence or absence of cancer in that organ or tissue.

For patients diagnosed with prostate cancer, injection of an antibody which specifically binds CSG can also have a therapeutic benefit. The antibody may exert its therapeutic effect alone. Alternatively, the antibody can be conjugated to a cytotoxic agent such as a drug, toxin or radionuclide to enhance its therapeutic effect. Drug monoclonal antibodies have been described in the art for example by Garnett and Baldwin, Cancer Research 1986 46:2407-2412. The use of toxins conjugated to monoclonal antibodies for the therapy of various cancers has also been described by Pastan et al. Cell 1986 47:641-648. Yttrium-90 labeled monoclonal antibodies have been described for maximization of dose delivered to the tumor while limiting toxicity to normal tissues (Goodwin and Meares Cancer Supplement 1997 80:2675-2680). Other cytotoxic radionuclides including, but not limited to Copper-67, Iodine-131 and Rhenium-186 can also be used for labeling of antibodies against CSG.

Antibodies which can be used in these *in vivo* methods include polyclonal, monoclonal and omniclonal antibodies and antibodies prepared via molecular biology techniques. Antibody fragments and aptamers and single-stranded oligonucleotides such as those derived from an *in vitro* evolution protocol referred to as SELEX and well known to those skilled in the art can also be used.

Small molecules predicted via computer imaging to specifically bind to regions of CSGs can also be designed and synthesized and tested for use in the imaging and treatment of prostate cancer. Further, libraries of molecules can be screened for potential anticancer agents by assessing the ability of the molecule to bind to CSGs identified herein. Molecules identified in the library as being capable of binding to CSG are key candidates for further evaluation for use in the treatment of prostate cancer.

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EXAMPLES

The present invention is further described by the following examples. These examples are provided solely to illustrate the invention by reference to specific embodiments.

- 5 These exemplifications, while illustrating certain aspects of the invention, do not portray the limitations or circumscribe the scope of the disclosed invention.

All examples outlined here were carried out using standard techniques, which are well known and routine to those
10 of skill in the art, except where otherwise described in detail. Routine molecular biology techniques of the following example can be carried out as described in standard laboratory manuals, such as Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory
15 Press, Cold Spring Harbor, N.Y. (1989).

Example 1: Identification of CSGs

Identification of CSGs were carried out by a systematic analysis of data in the LIFESEQ database available from Incyte Pharmaceuticals, Palo Alto, CA, using the data mining Cancer
20 Leads Automatic Search Package (CLASP) developed by diaDexus LLC, Santa Clara, CA.

The CLASP performs the following steps: selection of highly expressed organ specific genes based on the abundance level of the corresponding EST in the targeted organ versus
25 all the other organs; analysis of the expression level of each highly expressed organ specific genes in normal, tumor tissue, disease tissue and tissue libraries associated with tumor or disease; selection of the candidates demonstrating component ESTs were exclusively or more frequently found in tumor
30 libraries. The CLASP allows the identification of highly expressed organ and cancer specific genes. A final manual in depth evaluation is then performed to finalize the CSGs selection.

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Clones depicted in the following Table 1 are CSGs useful in diagnosing, monitoring, staging, imaging and treating prostate cancer.

Table 1: CSGs

5	Clone ID	Pro #	SEQ ID NO:
	3424528H1	Pro109	1,2
	578349H1	Pro112	3,4
	1794013H1	Pro111	5,6
	2189835H1	Pro115	7,8
10	3277219H1	Pro110	9,10
	1857415	Pro113	11,12
	1810463H1	Pro114	13,14
	zr65G11	Pro118	15,16
	2626135H1		17
15	zd46d08		18
	1712252H1		19
	784583H1		20

Example 2: Relative Quantitation of Gene Expression

Real-Time quantitative PCR with fluorescent Tagman probes is a quantitation detection system utilizing the 5'-3' nuclease activity of Taq DNA polymerase. The method uses an internal fluorescent oligonucleotide probe (Tagman) labeled with a 5' reporter dye and a downstream, 3' quencher dye. During PCR, the 5'-3' nuclease activity of Taq DNA polymerase releases the reporter, whose fluorescence can then be detected by the laser detector of the Model 7700 Sequence Detection System (PE Applied Biosystems, Foster City, CA, USA).

Amplification of an endogenous control is used to standardize the amount of sample RNA added to the reaction and normalize for Reverse Transcriptase (RT) efficiency. Either cyclophilin, glyceraldehyde-3-phosphate dehydrogenase (GAPDH), ATPase, or 18S ribosomal RNA (rRNA) is used as this endogenous

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control. To calculate relative quantitation between all the samples studied, the target RNA levels for one sample were used as the basis for comparative results (calibrator). Quantitation relative to the "calibrator" can be obtained using the standard curve method or the comparative method (User Bulletin #2: ABI PRISM 7700 Sequence Detection System).

The tissue distribution and the level of the target gene were evaluated for every sample in normal and cancer tissues. Total RNA was extracted from normal tissues, cancer tissues, and from cancers and the corresponding matched adjacent tissues. Subsequently, first strand cDNA was prepared with reverse transcriptase and the polymerase chain reaction was done using primers and Taqman probes specific to each target gene. The results were analyzed using the ABI PRISM 7700 Sequence Detector. The absolute numbers are relative levels of expression of the target gene in a particular tissue compared to the calibrator tissue.

Expression of Clone ID 3424528H1 (Pro109):

For the CSG Pro109, real-time quantitative PCR was performed using the following primers:

Forward Primer:

5'- ATCAGAACAAAGAGGCTGTGTC - 3' (SEQ ID NO:21)

Reverse Primer:

5'- ATCTCTAAAGCCCCAACCTTC - 3' (SEQ ID NO:22)

The absolute numbers depicted in Table 2 are relative levels of expression of the CSG referred to as Pro109 in 12 normal different tissues. All the values are compared to normal stomach (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

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Table 2: Relative Levels of CSG Prol9 Expression in Pooled Samples

Tissue	NORMAL
Colon	0.02
Endometrium	0.01
Kidney	0.48
Liver	14.83
Ovary	0.08
Pancreas	4.38
Prostate	11.24
Small Intestine	0.42
Spleen	0
Stomach	1
Testis	0.62
Uterus	0.02

The relative levels of expression in Table 2 show that with the exception of liver (14.83), Prol9 mRNA expression is higher (11.24) in prostate compared with all other normal tissues analyzed. Pancreas, with a relative expression level of 4.38, is the only other tissue expressing considerable mRNA for Prol9.

The absolute numbers in Table 2 were obtained analyzing pools of samples of a particular tissue from different individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 3.

The absolute numbers depicted in Table 3 are relative levels of expression of Prol9 in 28 pairs of matching samples and 4 unmatched samples. All the values are compared to normal stomach (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

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Table 3: Relative Levels of CSG Pro109 Expression in Individual Samples

Sample ID	Tissue	Cancer	Matching Normal Adjacent
Pro34B	Prostate 1	5.98	6.06
Pro65XB	Prostate 2	16.68	3.85
Pro69XB	Prostate 3	20.46	6.82
Pro78XB	Prostate 4	1.39	1.4
Pro101XB	Prostate 5	24.8	9.8
Pro12B	Prostate 6	9.1	0.2
Pro13XB	Prostate 7	0.5	9.7
Pro20XB	Prostate 8	13	12.5
Pro23B	Prostate 9	16.8	3
Ovr100050	Ovary 1	0.4	
Ovr1028	Ovary 2	1.9	
Ovr18GA	Ovary 3		0.1
Ovr206I	Ovary 4		0.1
Mam12X	Mammary Gland 1	13.5	1.4
Mam47XP	Mammary Gland 2	0.7	0.2
Lng47XQ	Lung 1	2.36	0.03
Lng60XL	Lung 2	7.39	0.2
Lng75XC	Lung 3	0.77	0.27
StoAC44	Stomach 1	0.05	1.19
StoAC93	Stomach 2	0.55	0.8
StoAC99	Stomach 3	0.12	3.04
ColAS43	Colon 1	16.11	0.07
ColAS45	Colon 2	0.11	0.08
ColAS46	Colon 3	4.99	0.4
Liv15XA	Liver 1	8.43	10.97
Liv42X	Liver 2	1.57	20.82

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Liv94XA	Liver 3	2.98	9.19
Pan77X	Pancreas 1	36	32
Pan82XP	Pancreas 2	0.09	7.09
Pan92X	Pancreas 3	0.7	0
Pan71XL	Pancreas 4	2.48	0.73
Pan10343	Pancreas 5	46	5.5

0 = Negative

In the analysis of matching samples, the higher levels of expression were in prostate, showing a high degree of tissue specificity for prostate tissue. Of all the samples different than prostate analyzed, only 4 cancer samples (the cancer sample mammary 1 with 13.5, colon 1 with 16.11, liver 1 with 8.43, and lung 2 with 7.39) showed an expression comparable to the mRNA expression in prostate. These results confirmed some degree of tissue specificity as obtained with the panel of normal pooled samples (Table 2).

Furthermore, the level of mRNA expression was compared in cancer samples and the isogenic normal adjacent tissue from the same individual. This comparison provides an indication of specificity for the cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 3 shows overexpression of Prol09 in 6 out of 9 primary prostate cancer tissues compared with their respective normal adjacents. Thus, overexpression in the cancer tissue was observed in 66.66% of the prostate matching samples tested (total of 9 prostate matching samples).

Altogether, the degree of tissue specificity, plus the mRNA overexpression in 66.66% of the primary prostate matching samples tested is indicative of Prol09 being a diagnostic marker for prostate cancer.

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Expression of Clone ID 578349H1 (Prol12):

For the CSG Prol12, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- TGCCGAAGAGGTTCACTGC - 3' (SEQ ID NO:23)

Reverse Primer

5'- GCCACAGTGGTACTGTCCAGAT - 3' (SEQ ID NO:24)

The absolute numbers depicted in Table 4 are relative levels of expression of the CSG Prol12 in 12 normal different tissues. All the values are compared to normal thymus (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 4: Relative Levels of CSG Prol12 Expression in Pooled Samples

Tissue	NORMAL
Brain	2.9
Heart	0.1
Kidney	0.2
Liver	0.2
Lung	7.7
Mammary	4.2
Muscle	0.1
Prostate	5.5
Small Intestine	1.8
Testis	1
Thymus	1
Uterus	21

The relative levels of expression in Table 4 show that Prol12 mRNA expression is the 3rd most highly expressed gene (after uterus and mammary) in the pool of normal prostate tissue compared to a total of 12 tissues analyzed. The absolute numbers in Table 4 were obtained analyzing pools of samples of a particular tissue from different individuals. These results demonstrate that Prol12 mRNA expression is specific for prostate thus indicating Prol12 to be a diagnostic marker for prostate disease especially cancer.

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Expression of Clone ID 1794013H1 (Prol11):

For the CSG Prol11, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- GCTGCAAGTTCTCCACATTGA - 3' (SEQ ID NO:25)

Reverse Primer

5'- CAGCCGCAGGTGAAACAC - 3' (SEQ ID NO:26)

The absolute numbers depicted in Table 5 are relative levels of expression of the CSG Prol11 in 12 normal different tissues. All the values are compared to normal testis (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 5: Relative Levels of CSG Prol11 Expression in Pooled Samples

Tissue	NORMAL
Brain	0.04
Heart	0
Kidney	0
Liver	0
Lung	0.05
Mammary	0.14
Muscle	5166.6
Prostate	1483.72
Small Intestine	0.33
Testis	1
Thymus	0.49
Uterus	0.07

The relative levels of expression in Table 5 show that Prol11 mRNA expression is extraordinarily high in the pool of normal prostate (1483.72) compared to all the other tissues analyzed with the exception of muscle (5166.6). These results demonstrate that Prol11 mRNA expression shows specificity for prostate and muscle.

The absolute numbers in Table 5 were obtained analyzing pools of samples of a particular tissue from different

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individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 6.

The absolute numbers depicted in Table 6 are relative levels of expression of Prol11 in 48 pairs of matching and 18 unmatched samples. All the values are compared to normal testis (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

Table 6: Relative Levels of CSG Prol11 Expression in Individual Samples

Sample ID	Tissue	Cancer	Matching Normal Adjacent
Pro101XB	Prostate 1	8.3	21.8
Pro12B	Prostate 2	2336	133
Pro13XB	Prostate 3	3.4	23
Pro20XB	Prostate 4	21.6	121.5
Pro23B	Prostate 5	19.4	3.7
Pro34B	Prostate 6	15	39
Pro65XB	Prostate 7	8	867
Pro69XB	Prostate 8	56	94
Pro78XB	Prostate 9	24	1515
Pro84XB	Prostate 10	119	15.35
Pro90XB	Prostate 11	8.08	112.2
Pro91XB	Prostate 12	0.88	51.8
ProC215	Prostate 13	0.3	
ProC234	Prostate 14	0.35	
ProC280	Prostate 15	436.5	
Pro109XB	Prostate 16	3.43	265
Pro110	Prostate 17	18.2	8.73

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	Pro125XB	Prostate 18	0.34	186
	Pro326	Prostate 19	1392	110
	Pro10R	Prostate 20 (prostatitis)	0.5	
	Pro20R	Prostate 21 (prostatitis)	24.1	
5	Pro258	Prostate 22 (BPH)	4610	
	Pro263C	Prostate 23 (BPH)	0	
	Pro267A	Prostate 24 (BPH)	1.46	
	Pro271A	Prostate 25 (BPH)	0	
	Pro460Z	Prostate 26 (BPH)	1.47	
10	ProC032	Prostate 27 (BPH)	14.4	
	Tst39X	Testis 1	0	0
	Bld32XK	Bladder 1	0.44	0.41
	Bld46XK	Bladder 2	0	0
	Bld66X	Bladder 3	0	0
15	BldTR14	Bladder 4	0	0
	Kid106XD	Kidney 1	0	0
	Kid107XD	Kidney 2	0	0
	Kid109XD	Kidney 3	0	0
	Pan10343	Pancreas 1	0	0
20	Pan71XL	Pancreas 2	0	0
	Pan77X	Pancreas 3	0	0
	Liv15XA	Liver 1	0	0
	Liv42X	Liver 2	0	0
	ClnAS43	Colon 1	0	0
25	ClnAS45	Colon 2	0	0
	ClnAS46	Colon 3	0	0
	ClnAS67	Colon 4	0	0
	ClnAC19	Colon 5	0	0
	ClnAS12	Colon 6	0	0

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	SmI21XA	Small Intestine 1	0	0
	SmIH89	Small Intestine 2	0	0
	Lng47XQ	Lung 1	0.7	0
	Lng60XL	Lung 2	0	0
5	Lng75XC	Lung 3	0	0
	Lng90X	Lung 4	0	0
	Mam12X	Mammary Gland 1	0	1.4
	Mam59X	Mammary Gland 2	0.2	0
	MamA06X	Mammary Gland 3	0	0
10	MamS127	Mammary Gland 4	0	0
	Mam162X	Mammary Gland 5	0	0
	Mam42DN	Mammary Gland 6	0	0
	Ovr103X	Ovary 1	0.14	0
	Ovr10050	Ovary 2	0.2	
15	Ovr1028	Ovary 3	0	
	Ovr10400	Ovary 4	0.2	
	Ovr18GA	Ovary 5		0
	Ovr206I	Ovary 6		0
	Ovr20GA	Ovary 7		0.2
20	Ovr25GA	Ovary 8		0

0= Negative

In the analysis of matching samples, the higher levels of expression were in prostate showing a high degree of tissue specificity for prostate. These results confirm the tissue specificity results obtained with normal pooled samples (Table 5).

Furthermore, the level of mRNA expression in cancer samples and the isogenic normal adjacent tissue from the same individual were compared. This comparison provides an indication of specificity for cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 6 shows overexpression of Prol11 in 5 out

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of 16 primary prostate cancer samples compared with their respective normal adjacent (prostate samples 2, 5, 10, 17, and 19). Similar expression levels were observed in 3 unmatched prostate cancers (prostate samples 13, 14, 15), 2 prostatitis (prostate samples 20, 21), and 6 benign prostatic hyperplasia samples (prostate samples 22 through 27). Thus, there is overexpression in the cancer tissue of 31.25% of the prostate matching samples tested (total of 16 prostate matching samples).

10 Altogether, the high level of tissue specificity, plus the mRNA overexpression in 31.25% of the prostate matching samples tested are indicative of Prol11 being a diagnostic marker for prostate cancer.

Expression of Clone ID 2189835H1 (Prol115):

15 For the CSG Prol115, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- TGGCTTTGAACTCAGGGTCA - 3' (SEQ ID NO:27)

Reverse Primer

20 5'- CGGATGCACCTCGTAGACAG - 3' (SEQ ID NO:28)

The absolute numbers depicted in Table 7 are relative levels of expression of the CSG Prol115 in 12 normal different tissues. All the values are compared to normal thymus (calibrator). These RNA samples are commercially available
25 pools, originated by pooling samples of a particular tissue from different individuals.

Table 7: Relative Levels of CSG Prol115 Expression in Pooled Samples

Tissue	NORMAL
Brain	0.016
Heart	0.002
Kidney	8.08
Liver	2.20
Lung	112.99

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Mammary	29.45
Muscle	0.05
Prostate	337.79
Small Intestine	7.54
Testis	1.48
Thymus	1
Uterus	1.4

The relative levels of expression in Table 7 show that Prol15 mRNA expression is higher (337.79) in prostate compared with all the other normal tissues analyzed. Lung, with a relative expression level of 112.99, and mammary (29.446) are the other tissues expressing moderate levels of mRNA for Prol15. These results establish Prol15 mRNA expression to be highly specific for prostate.

The absolute numbers in Table 7 were obtained analyzing pools of samples of a particular tissue from different individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 8.

The absolute numbers depicted in Table 8 are relative levels of expression of Prol15 in 17 pairs of matching and 21 unmatched samples. All the values are compared to normal thymus (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

Table 8: Relative Levels of CSG Prol15 Expression in Individual Samples

Sample ID	Tissue	Cancer	Matching Normal Adjacent
Prol12B	Prostate 1	1475.9	190.3
ProC234	Prostate 2	169.61	
Pro109XB	Prostate 3		639.53
Pro101XB	Prostate 4	1985.2	2882.9

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	Pro13XB	Prostate 5	34.9	13.9
	Pro215	Prostate 6	525.59	
	Pro125XB	Prostate 7		556.05
	Pro23B	Prostate 8	1891.4	1118.6
5	ProC280	Prostate 9	454.3	
	Pro20XB	Prostate 10	1332.6	
	Pro34B	Prostate 11		362.91
	Pro65XB	Prostate 12		135.06
	Pro69XB	Prostate 13		179.67
10	Pro10R	Prostate 14 (prostatitis)	143.82	
	Pro20R	Prostate 15 (prostatitis)	397.79	
	Pro258	Prostate 16 (BPH)	216.6	
	Pro263C	Prostate 17 (BPH)	601.25	
	Pro267A	Prostate 18 (BPH)	200.28	
15	Pro271A	Prostate 19 (BPH)	111.43	
	Pro460Z	Prostate 20 (BPH)	53.84	
	ProC032	Prostate 21 (BPH)	56.94	
	SmI21XA	Small Intestine 1	28.8	29.9
	SmIH89	Small Intestine 2	70.8	348.5
20	ClnAC19	Colon 1	22.73	446.47
	ClnAS12	Colon 2	116.97	493.18
	Kid106XD	Kidney 1	86.13	41.14
	Kid107XD	Kidney 2	0.26	35.14
	Lng47XQ	Lung 1	5.13	20.98
25	Lng60XL	Lung 2	13.93	114.78
	Lng75XC	Lung 3	16.47	53.79
	Mam12X	Mammary Gland 1	6.25	10.75
	Mam162X	Mammary Gland 2	1.84	2.54
	Mam42DN	Mammary Gland 3	23.08	35.51

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	Ovr10050	Ovary 1	0.9	
	Ovr1028	Ovary 2	261.4	
	Ovr103X	Ovary 3	7	0.1
	Ovr20GA	Ovary 4		0
5	Ovr25GA	Ovary 5		0

0 = Negative

Higher levels of expression were seen in prostate, showing a high degree of tissue specificity for prostate tissue. Of all the analyzed samples different from prostate, only two cancer samples (colon 2 with 116.97 and ovary 2 with 261.4), and 5 normal adjacent tissue samples (small intestine 2, colon 1, colon 2, kidney 1, and lung 2), showed an expression comparable to the mRNA expression in prostate. These results confirmed the tissue specificity results obtained with the panel of normal pooled samples (Table 7).

Furthermore, the levels of mRNA expression in cancer samples and the isogenic normal adjacent tissue from the same individual were compared. This comparison provides an indication of specificity for the cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 8 shows higher expression of Prol15 in 3 out of 4 matched prostate cancer tissues (prostate samples 1, 5 & 8).

Altogether, the high level of tissue specificity, plus the higher expression in 75% of the prostate matching samples tested, are indicative of Prol15 being a diagnostic marker for prostate cancer.

Expression of Clone ID 3277219H1 (Prol10):

For the CSG Prol10, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- CGGCAACCTGGTAGTGAGTG - 3' (SEQ ID NO:29)

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Reverse Primer

5'- CGCAGCTCCTTGTAACCTCAG - 3' (SEQ ID NO:30)

The absolute numbers depicted in Table 9 are relative levels of expression of the CSG Prol10 in 12 normal different tissues. All the values are compared to normal small intestine (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 9: Relative Levels of CSG Prol10 Expression in Pooled Samples

Tissue	NORMAL
Brain	6.61
Heart	0.7
Kidney	0.74
Liver	7.94
Lung	11.88
Mammary	22.78
Muscle	6.77
Prostate	3.01
Small Intestine	1
Testis	2.58
Thymus	13.74
Uterus	2.61

The relative levels of expression in Table 9 show that Prol10 mRNA expression is not as high in normal prostate (3.01) compared with all the other normal tissues analyzed.

The absolute numbers in Table 9 were obtained analyzing pools of samples of a particular tissue from different individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 10.

The absolute numbers depicted in Table 10 are relative levels of expression of Prol10 in 33 pairs of matching samples. All the values are compared to normal small intestine (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from

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the normal adjacent sample for that same tissue from the same individual.

Table 10: Relative Levels of CSG Prol10 Expression in Individual Samples

	Sample ID	Tissue	Cancer	Matching Normal Adjacent
5	Pro12B	Prostate 1	11.8	0.3
	Pro78XB	Prostate 2	14.3	6.3
	Pro101XB	Prostate 3	33.2	10.7
	Pro13XB	Prostate 4	0.3	0.4
10	Pro23XB	Prostate 5	25.5	14.4
	Pro20XB	Prostate 6	43.3	4
	Pro34XB	Prostate 7	31.8	18.7
	Pro65XB	Prostate 8	26.9	3.4
	Pro69XB	Prostate 9	12.5	7
15	Lng75XC	Lung 1	1.9	3
	Lng90X	Lung 2	5.5	0.5
	LngAC11	Lung 3	9.3	9.7
	LngAC32	Lung 4	11.2	2.2
	Lng47XQ	Lung 5	11.3	0.3
20	Lng60XL	Lung 6	29.1	6.8
	Mam12B	Mammary Gland 1	19.8	0
	Mam603X	Mammary Gland 2	13.7	0
	Mam82XI	Mammary Gland 3	73.5	0
	MamA04	Mammary Gland 4	0	24.6
25	MamB011X	Mammary Gland 5	17.4	2
	MamC012	Mammary Gland 6	0	12.8
	MamC034	Mammary Gland 7	0	61
	Mam12X	Mammary Gland 8	14	2.2
	Mam59X	Mammary Gland 9	33	2.2

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MamA06X	Mammary Gland 10	16.4	0.8
Liv15XA	Liver 1	4.7	0.6
Liv42X	Liver 2	7.5	2.6
Liv94XA	Liver 3	0.4	1.4
ClnAS43	Colon 1	52.9	1.4
ClnAS45	Colon 2	2.1	0.8
ClnAS46	Colon 3	39.8	3.7
SmI21X	Small Intestine 1	0.9	0.1
SmIH89	Small Intestine 2	5.8	0.9

0 = Negative

The levels of mRNA expression in cancer samples and the isogenic normal adjacent tissue from the same individual were compared. This comparison provides an indication of specificity for the cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 10 shows overexpression of Prol10 in 8 of the 9 primary prostate cancer tissues compared with their respective normal adjacent (except prostate 4). Thus, there was overexpression in 88.88% of the cancer prostate tissue as compared to the prostate matching samples tested (total of 9 prostate matching samples).

Although not tissue specific, Prol10 mRNA expression is upregulated in prostate cancer tissues. The mRNA overexpression in 88.88% of the primary prostate matching cancer samples tested is indicative of Prol10 being a diagnostic marker for prostate cancer. Prol10 also showed overexpression in several other cancers tested including small intestine, colon, liver, mammary and lung (see Table 10). Accordingly Prol10 may be a diagnostic marker for other types of cancer as well.

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Expression of Clone ID 1857415; Gene ID 346880 (Prol13):

For the CSG Prol13, real-time quantitative PCR was performed using the following primers:

Forward Primer

5' - CGGGAACCTACCAGCCTATG - 3' (SEQ ID NO:31)

Reverse Primer

5' - CAGGCAACAGGGAGTCATGT - 3' (SEQ ID NO:32)

The absolute numbers depicted in Table 11 are relative levels of expression of the CSG Prol13 in 12 normal different tissues. All the values are compared to normal thymus (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 11: Relative Levels of CSG Prol13 Expression in Pooled Samples

Tissue	NORMAL
Brain	0.03
Heart	0
Kidney	0.01
Liver	0
Lung	0
Mammary Gland	0
Muscle	0.04
Prostate	489.44
Small Intestine	0.02
Testis	0.35
Thymus	1
Uterus	0.13

The relative levels of expression in Table 11 show that Prol13 mRNA expression is higher (489.44) in prostate compared with all the other normal tissues analyzed. Testis, with a relative expression level of 0.35, uterus (0.13), thymus (1.0), kidney (0.01) and brain (0.03) were among the other tissues expressing lower mRNA levels for Prol13. These results establish that Prol13 mRNA expression is highly specific for prostate.

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The absolute numbers in Table 11 were obtained analyzing pools of samples of a particular tissue from different individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 12.

The absolute numbers depicted in Table 12 are relative levels of expression of Prol13 in 78 pairs of matching and 25 unmatched tissue samples. All the values are compared to normal thymus (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual. In cancers (for example, ovary) where it was not possible to obtain normal adjacent samples from the same individual, samples from a different normal individual were analyzed.

Table 12: Relative Levels of CSG Prol13 Expression in Individual Samples

Sample ID	Tissue	Cancer	Matched or Unmatched Normal Adjacent
Pro780B/781B	Prostate 1	375.58	446.29
Pro1291B/1292B	Prostate 2	1060	31
Pro139B96/140B96	Prostate 3	41	32
Pro209B96/210B96	Prostate 4	505	255
Pro1256B/1257B	Prostate 5	165.79	141.63
Pro1293B/1294B	Prostate 6	1613.7	874.61
Pro694B/695B	Prostate 7	458.6	142.21
Pro1012B/1013B	Prostate 8	1520	864
Pro1222B/1223B	Prostate 9	939	530
Pro845B/846B	Prostate 10	1552.4	374.6
Pro1094B/1095B	Prostate 11	278.37	135.89
Pro650B/651B	Prostate 12	532.81	640.85

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	Pro902B/903B	Prostate 13	609.05	415.86
	Pro916B/917B	Prostate 14	699.42	401.24
	Pro9821110A/110B	Prostate 15	156	487.8
	ProS9821326A/26B	Prostate 16	744.4	472.8
5	Pro9407c215	Prostate 17	1389.2	
	Pro9407c234	Prostate 18	305.5	
	Pro9407c280A	Prostate 19	894.5	
	Pro9409C010R	Prostate 20 (prostatitis)	269.7	
	Pro9404C120R	Prostate 21 (prostatitis)	299.2	
10	Pro1000258	Prostate 22 (BPH)	149.6	
	Pro4001263C	Prostate 23 (BPH)	576	
	Pro4001267A	Prostate 24 (BPH)	132.1	
	Pro9411C032	Prostate 25 (BPH)	118.2	
	Pro4001460Z	Prostate 26 (BPH)	276.3	
15	Pro4001271A	Prostate 27 (BPH)	58.7	
	Kid1064D/65D	Kidney 1	0	0.1
	Kid1079D/1080D	Kidney 2	0.3	0.02
	Kid1097D/1098D	Kidney 3	35.14	0.32
	Kid1024D/1025D	Kidney 4	1.31	0
20	Kid1183D/1184D	Kidney 5	24.79	0
	Kid1242D/1243D	Kidney 6	0	0
	Bld469K	Bladder 1		2.88
	Bld467K/468K	Bladder 2	2.65	
	Bld327K/328K	Bladder 3	0	4.05
25	Bld470K	Bladder 4		1.64
	Bld665T/664T	Bladder 5	0.21	1.99

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	Bld1496K/1497K	Bladder 6	13.55	1.14
	Bld1721K/1722K	Bladder 7	120.16	1.34
	Tst239X/240X	Testis 1	31.5	0.73
	TstS9820647A/47B	Testis 2	15.7	0
5	TstS9820663A/663B	Testis 3	72	1.4
	SknS9821248A/248B	Skin 1	1.8	0.5
	SknS99448A/448B	Skin 2	251.6	0
	Skn99816A/816B	Skin 3	33	0.7
	Sto4004864A4/B4	Stomach 1	14.12	0
10	Sto4004509A3/B1	Stomach 2	40.74	39
	SmI9807A212A/213A	Small Intestine 1	0.1	0
	SmI9802H008/H009	Small Intestine 2	5.8	0.1
	Cln9608B012/B011	Colon 1	4.5	0
	Cln9709c074ra/073ra	Colon 2	65.8	3.1
15	Cln4004709A1/709B1	Colon 3	1.1	0.9
	Cln9405C199/C200	Colon 4	34.76	0.73
	Cln9707c004gb/006ga	Colon 5	90.26	0.96
	Cln96-09-B004/B003	Colon 6	17.9	20.64
	Cln9612B006/B005	Colon 7	17.56	0.3
20	Cln9705F002D/F001C	Colon 8	21.39	0
	ClnCXGA	Colon 9	429.14	142.69
	Pan10343a	Pancreas 1	0	0
	Pan776P/777P	Pancreas 2	0	0.15
	Pan9210/9220	Pancreas 3	7.36	0
25	Pan714L/715L	Pancreas 4	13.57	0.11
	Pan824P/825P	Pancreas 5	0	0
	Lng476Q/477Q	Lung 1	0	0
	Lng605L/606L	Lung 2	0	0.1
	Lng11145B/11145C	Lung 3	85.9	0

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Lng0008632A/32B	Lung 4	23.85	0
Lng750C/751C	Lung 5	0.32	0.25
Lng8890A/8890B	Lung 6	10.63	0
Lng8926A/8926B	Lung 7	15.37	0
Lng0010239A/39B	Lung 8	26.17	0
Lng9502C109R/110R	Lung 9	0.68	0
LngS9821944a/44b	Lung 10	0	0
Mam00042D01/42N01	Mammary Gland 1	8.5	0
Mam59XC	Mammary Gland 2	61.07	0
Mam9706A066G/67C	Mammary Gland 3	4.84	0
Mam14153a1C	Mammary Gland 4	9.72	6.99
Mam1620F/1621F	Mammary Gland 5	0.91	0
Mam00014D05	Mammary Gland 6	2.45	0
End10479B/D	Endometrium 1	133.43	1.12
End9705A125A/126A	Endometrium 2	0	0.39
End9704C281A/282A	Endometrium 3	23.5	1.56
End680c97/681c97	Endometrium 4	88.89	79.02
Utr13590/13580	Uterus 1	0.2	0
Utr850U/851U	Uterus 2	0	0
Utr14170/14180	Uterus 3	14	0.4
Utr233U96/234U96	Uterus 4	8.65	4.64
CvxVNM00052D01/52N01	Cervix 1	0.82	77.15
CvxVNM00083D01/83N01	Cervix 2	0.78	221.48
CvxND00023D01/23N01	Cervix 3	3.25	15.22
Ovr10370/10380	Ovary 1	0.1	0
Ovr10050	Ovary 2	18.96	
Ovr1028	Ovary 3	0	
Ovr14638A1C	Ovary 4	3.2	
Ovr14603A1D	Ovary 5	882.3	
Ovr7730	Ovary 6	0	

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Ovr9702C018GA	Ovary 7	0.15
Ovr206I	Ovary 8	0
Ovr9702C020GA	Ovary 9	0
Ovr9702C025GA	Ovary 10	0
Ovr9701C035GA	Ovary 11	0.07
Ovr9701C050GB	Ovary 12	0.58

0 = Negative

In the analysis of matching samples, the higher levels of expression were in prostate, showing a high degree of tissue specificity for prostate tissue. In addition to the higher expression levels in prostate cancer samples, Prol13 expression was found to be either induced (where not expressed in normal adjacent tissues) or somewhat upregulated in several other cancers. However, the relative expression and the fold increase in prostate cancer samples far exceeds that in other cancer tissues and is highly significant.

Furthermore, the levels of mRNA expression in cancer samples and the isogenic normal adjacent tissue from the same individual were compared. This comparison provides an indication of specificity for the cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 12 shows overexpression of Prol13 in 13 out of 16 primary prostate cancer tissues compared with their respective normal adjacent (prostate samples 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 13, 14, 16). Thus, there was overexpression in the cancer tissue for 81.25% of the prostate matching samples tested. The median for the level of expression in prostate cancer tissue samples is 609, whereas the median for all other cancers is only 7.93, with the exception of one colon sample, colon 9, whose expression was similar to that found in prostate cancer tissues.

Altogether, the high level of tissue specificity, plus the mRNA overexpression in 81.25% of the primary prostate matching samples tested are indicative of Prol13 being a

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diagnostic marker for prostate cancer. Expression was also found to be higher in other cancer tissues compared with their respective normal adjacent tissues (kidney, bladder, testis, skin, stomach, small intestine, colon, pancreas, lung, mammary, endometrium, uterus, and ovary) thus indicating Prol13 to be a pan cancer marker.

Expression of Clone ID 1810463H1 (Prol14):

For the CSG Prol14, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- TGGGCATCTGGGTGTCAA - 3' (SEQ ID NO:33)

Reverse Primer

5'- CGGCTGCGATGAGGAAGTA - 3' (SEQ ID NO:34)

The absolute numbers depicted in Table 13 are relative levels of expression of the CSG Prol14 in 12 normal different tissues. All the values are compared to normal muscle (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 13: Relative Levels of CSG Prol14 Expression in Pooled Samples

Tissue	NORMAL
Brain	9.7
Heart	0.7
Kidney	414.4
Liver	4
Lung	882.2
Mammary	44
Muscle	1
Prostate	1951
Small Intestine	22
Testis	367.1
Thymus	25.8
Uterus	139.6

The relative levels of expression in Table 13 show that Prol14 mRNA expression is higher (1951) in prostate compared with all the other normal tissues analyzed. Lung, with a relative

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expression level of 882.2, kidney 414.4, testis 367.1 and uterus 139.6, are the other tissues expressing higher levels of mRNA for Proll14. These results establish Proll14 mRNA expression to be more specific for prostate than other tissues examined.

The high level of tissue specificity is indicative of Proll14 being a diagnostic marker for diseases of the prostate, especially cancer.

Expression of Clone ID zr65g11 (Proll18):

For the CSG Proll18, real-time quantitative PCR was performed using the following primers:

Forward Primer

5'- GCCCATCTCCTGCTTCTTTAGT - 3' (SEQ ID NO:35)

Reverse Primer

5'- CGTGGAGATGGCTCTGATGTA - 3' (SEQ ID NO:36)

The absolute numbers depicted in Table 14 are relative levels of expression of the CSG Proll18 in 12 normal different tissues. All the values are compared to normal kidney (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 14: Relative Levels of CSG Proll18 Expression in Pooled Samples

Tissue	NORMAL
Colon	0.87
Endometrium	19282
Kidney	1
Liver	0
Ovary	86.22
Pancreas	0
Prostate	962.1
Small Intestine	0
Spleen	0.75
Stomach	0.54
Testis	343.7
Uterus	1064

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The relative levels of expression in Table 4 show that Prol18 mRNA expression is the 3rd highest in prostate (962.1) next to endometrium (19282) and uterus (1064), which are female-specific tissues. Testis, with a relative expression level of 343.7 is the only other male tissue expressing moderate levels of mRNA for Prol18. These results establish Prol18 mRNA expression to be highly specific for reproductive tissues including the prostate.

The absolute numbers in Table 14 were obtained analyzing pools of samples of a particular tissue from different individuals. They cannot be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 15.

The absolute numbers depicted in Table 15 are relative levels of expression of Prol18 in 59 pairs of matching and 21 unmatched samples. All the values are compared to normal kidney (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

Table 15: Relative Levels of CSG Prol18 Expression in Individual Samples

Sample ID	Tissue	Cancer	Matching Normal Adjacent
Prol2B	Prostate 1	41700.7	22242.83
ProC234	Prostate 2	40087	
Pro78XB	Prostate 3	4075.6	7066.7
Pro109XB	Prostate 4	334.4	777.2
Pro84XB	Prostate 5	11684	58290
Pro101XB	Prostate 6	21474.13	100720.8
Pro91X	Prostate 7	14849	33717
Pro13XB	Prostate 8	202.57	146.91

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	ProC215	Prostate 9	73243	
	Pro125XB	Prostate 10	629.6	521.4
	Pro23B	Prostate 11	157532.6	110654.4
	Pro90XB	Prostate 12	2317	64134
5	ProC280	Prostate 13	42020	
	Pro20XB	Prostate 14	2909.31	
	Pro34B	Prostate 15	29610	23264
	Pro110	Prostate 16	13354	30991
	Pro65XB	Prostate 17	10126	11270
10	Pro69XB	Prostate 18		2671.42
	Pro326	Prostate 19	9962.3	19231
	Pro10R	Prostate 20 (prostatitis)	27355	
	Pro20R	Prostate 21 (prostatitis)	21081	
	Pro258	Prostate 22 (BPH)	79916.32	
15	Pro263C	Prostate 23 (BPH)	108924.5	
	Pro267A	Prostate 24 (BPH)	92910.22	
	Pro271A	Prostate 25 (BPH)	57004.4	
	Pro460Z	Prostate 26 (BPH)	57449.23	
	ProC032	Prostate 27 (BPH)	45781.44	
20	Kid106XD	Kidney 1	3.08	217.36
	Kid107XD	Kidney 2	0	38.36
	Kid109XD	Kidney 3	0	123.5
	Kid10XD	Kidney 4	17.69	67.8
	Kid11XD	Kidney 5	16.74	360.8
25	Kid124D	Kidney 6	0	167.4
	Bld32XK	Bladder 1	0	0
	Bld47K	Bladder 2		36.38
	Bld66X	Bladder 3	0	4.52
	BldTR14	Bladder 4	0	12.17

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	End3AX	Endometrium 2	27825	65839
	End4XA	Endometrium 3	10.3	15935
	Utr1410	Uterus 1	18885	18116
	Utr23XU	Uterus 2	3358	7674
5	CvxKS52	Cervix 1	0	0
	CvxKS83	Cervix 2	0	0
	Ovr10050	Ovary 1	72.86	
	Ovr1028	Ovary 2	0	
	Ovr638A	Ovary 3	0	
10	Ovr63A	Ovary 4	90.88	
	Ovr7730	Ovary 5	1.21	
	Ovr10400	Ovary 6	5.08	
	Ovr1050	Ovary 7	0	
	Ovr1118	Ovary 8	7.41	
15	Ovr103X	Ovary 9		32.78
	Ovr20GA	Ovary 10		0
	Ovr25GA	Ovary 11		1173.83
	Ovr35GA	Ovary 12		313.4
	Ovr50GB	Ovary 13		823.1
20	Ovr18GA	Ovary 14		40.6
	Ovr206I	Ovary 15		1264
	Ovr230A	Ovary 16		1285

0 = Negative

In the analysis of matching samples, the higher levels of expression were in prostate, endometrium, testis, and ovary showing a high degree of tissue specificity for reproductive tissues. These results confirmed the tissue specificity results obtained with the panel of normal pooled samples (Table 14).

Furthermore, the levels of mRNA expression in cancer samples and the isogenic normal adjacent tissue from the same individual were compared. This comparison provides an

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indication of specificity for the cancer (e.g. higher levels of mRNA expression in the cancer sample compared to the normal adjacent). Table 15 shows overexpression of Prol18 in 5 out of 14 primary prostate cancer tissues (prostate samples 1, 8, 10, 11, 15) compared with their respective normal adjacent. Thus, there was overexpression in the cancer tissue for 35.71% of the prostate matching samples tested (total of 14 prostate matching samples). Expression of Prol18 was similarly higher in 3 unmatched cancer tissues (prostate samples 9, 13, 14), 2 prostatitis (prostate samples 20, 21), and 6 benign hyperplasia tissues (prostate samples 22 through 27).

Altogether, the high level of tissue specificity, plus the mRNA overexpression in 35.71% of the primary prostate matching samples tested are indicative of Prol18 being a diagnostic marker for prostate cancer.

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What is claimed is:

1. A method for diagnosing the presence of prostate cancer in a patient comprising:

(a) determining levels of CSG in cells, tissues or bodily
5 fluids in a patient; and

(b) comparing the determined levels of CSG with levels of CSG in cells, tissues or bodily fluids from a normal human control, wherein a change in determined levels of CSG in said patient versus normal human control is associated with the presence of prostate cancer.

2. A method of diagnosing metastases of prostate cancer in a patient comprising:

(a) identifying a patient having prostate cancer that is not known to have metastasized;

15 (b) determining CSG levels in a sample of cells, tissues,
or bodily fluid from said patient; and

(c) comparing the determined CSG levels with levels of CSG in cells, tissue, or bodily fluid of a normal human control, wherein an increase in determined CSG levels in the 20 patient versus the normal human control is associated with a cancer which has metastasized.

3. A method of staging prostate cancer in a patient having prostate cancer comprising:

(a) identifying a patient having prostate cancer;

25 (b) determining CSG levels in a sample of cells, tissue, or bodily fluid from said patient; and

(c) comparing determined CSG levels with levels of CSG in cells, tissues, or bodily fluid of a normal human control wherein an increase in determined CSG levels in said patient 30 versus the normal human control is associated with a cancer which is progressing and a decrease in the determined CSG levels is associated with a cancer which is regressing or in remission.

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4. A method of monitoring prostate cancer in a patient for the onset of metastasis comprising:

(a) identifying a patient having prostate cancer that is not known to have metastasized;

5 (b) periodically determining levels of CSG in samples of cells, tissues, or bodily fluid from said patient; and

(c) comparing the periodically determined CSG levels with levels of CSG in cells, tissues, or bodily fluid of a normal human control, wherein an increase in any one of the
10 periodically determined CSG levels in the patient versus the normal human control is associated with a cancer which has metastasized.

5. A method of monitoring a change in stage of prostate cancer in a patient comprising:

15 (a) identifying a patient having prostate cancer;

(b) periodically determining levels of CSG in cells, tissues, or bodily fluid from said patient; and

(c) comparing the periodically determined CSG levels with levels of CSG in cells, tissues, or bodily fluid of a normal
20 human control, wherein an increase in any one of the periodically determined CSG levels in the patient versus the normal human control is associated with a cancer which is progressing in stage and a decrease is associated with a cancer which is regressing in stage or in remission.

25 6. A method of identifying potential therapeutic agents for use in imaging and treating prostate cancer comprising screening molecules for an ability to bind to CSG wherein the ability of a molecule to bind to CSG is indicative of the molecule being useful in imaging and treating prostate cancer.

30 7. The method of claim 1, 2, 3, 4, 5 or 6 wherein the CSG comprises SEQ ID NO:1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12,

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13, 14, 15, 16, 17, 18, 19 or 20 or a polypeptide encoded thereby.

8. An antibody which specifically binds CSG.

9. A method of imaging prostate cancer in a patient
5 comprising administering to the patient an antibody of claim 8.

10. The method of claim 9 wherein said antibody is labeled with paramagnetic ions or a radioisotope.

11. A method of treating prostate cancer in a patient
10 comprising administering to the patient an antibody of claim 7.

12. The method of claim 11 wherein the antibody is conjugated to a cytotoxic agent.

Docket No.
DEX-0196

Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

Method of Diagnosing, Monitoring, Staging, Imaging and Treating Prostate Cancer

the specification of which

(check one)

☐ is attached hereto.

☒ was filed on 19 October 1999 as United States Application No. or PCT International

Application Number PCT/US99/24331

and was amended on _____

(if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) or Section 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Not Claimed

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

60/104,737	19 October 1998
(Application Serial No.)	(Filing Date)
_____	_____
(Application Serial No.)	(Filing Date)
_____	_____
(Application Serial No.)	(Filing Date)

I hereby claim the benefit under 35 U. S. C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, C. F. R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)
_____	_____	_____
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)
_____	_____	_____
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (list name and registration number)



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